Autonomic Diabetic Neuropathy Impairs Glucose and Dipeptidyl Peptidase 4 Inhibitor-Regulated Glucagon Concentration in Type 1 Diabetic Patients

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Abstract

Background: Dipeptidyl peptidase 4 inhibitors (DPP4i) could exert their glucagonostatic action through a functional autonomic nervous system independently from insulin secretion. We explored this hypothesis.

Methods: We studied C-peptide negative type 1 diabetic patients (T1D) with (AN+) or without (AN-) autonomic neuropathy. Plasma glucagon, active and total glucagon-like peptide-1 (GLP-1), GIP, pancreatic polypeptide (PP), and glucose concentrations were quantified over 180 minutes following a meal test. Furthermore, to increase the plasma concentration of GLP-1 between groups and study its impact on glucagon secretion, 50 mg of vildagliptin and a second meal test were administered to the same patients.

Results: The plasma concentration of glucagon was higher in AN+ patients than in AN- patients, which was associated with lower PP and active GLP-1 plasma concentrations. This first set of data suggest that AN, presumably involving parasympathetic activity, results in loss of glucose-regulated glucagon secretion. After DPP4i treatment, AN+ patients lost the ability to suppress plasma glucagon, and the low plasma PP responses were not restored.

Conclusions: We here show that a functional autonomic nervous system is required for the proper control of glucagon secretion. The mechanism is insulin-independent. The glucagonostatic action of DPP4i also requires this mechanism.

Keywords: Neuropathy; Gut brain axis; Incretins; GLP-1; Type 1 diabetes

Introduction

Elevated or inappropriately high concentrations of glucagon in plasma, relative to plasma glucose and insulin concentrations, characterize type 1 (T1DM) and type 2 diabetic (T2DM) patients. Glucagon-like peptide-1 (GLP-1) exerts a glucagonostatic effect in a glucose-dependent manner meaning that there is no inhibition of glucagon responses to hypoglycemia, which means that GLP-1 does not impair the role of glucagon in the defense against hypoglycemia when it is used therapeutically. This might suggest that the inhibitory mechanism could involve insulin secretion, but it is well established that GLP-1 as well as dipeptidyl peptidase 4 inhibitors (DPP4i) lowers glucagon secretion also in T1D patients with no residual beta cell function, indicating that the effect of GLP-1 on glucagon secretion is mediated, at least in this situation, by a mechanism independent from insulin secretion [1-5]. Such glucose-dependent and insulin independent effects could involve neural mechanisms, since the plasma glucagon concentration is under the control of glucose-sensitive neural units mostly located within the brain [6, 7]. These units appear to be able to detect the glycemic variations and signal to the pancreatic alpha cells to enhance or reduce secretion. Some of the molecular mechanisms have been elucidated and appear to involve the glucose transporter GLUT2 [7], the inward rectifying potassium channel Kir 6.2 [8, 9] and perhaps the brain GLP-1 receptor [10]. Regarding GLP-1, an endocrine regulation directly on the pancreatic alpha cells for the control of glucagon secretion would seem incompatible with the rapid degradation of GLP-1 by the DPP4 which means that very little GLP-1 reaches the pancreas in the intact form [11]. Furthermore, whether or not the GLP-1 receptor on pancreatic alpha cell is responsible for the glucagonostatic effect of GLP-1 remains highly controversial. For similar reasons, a mechanism involving stimulation by GLP-1 of somatostatin secreted from neighboring delta cells,
which can be demonstrated in the perfused rat pancreas [12], may be relevant only for GLP-1 agonist administered in high doses. Together, this has called for other, non-endocrine, hypotheses. Our hypothesis is that enteric GLP-1 might activate a gut-to-brain-to-pancreatic alpha cell neuronal axis to control glucagon secretion during a meal. This hypothesis might help to explain the glucagonostatic effect of the DPP4i, since they are known to augment the concentration of active GLP-1 in the blood of the hepatoportal vein [11, 13-15]. This hypothesis was initially generated more than a decade ago during experiments which showed that GLP-1 and the GLP-1 receptor trigger the vagus nerve activity in response to glucose [16-18] and subsequently confirmed by independent researchers [19-21], also in humans [22]. This pathway might play an important role in the neural control of glucagon secretion [23-26]. We recently further characterized this gut-to-brain-to-pancreas axis by using low dose oral DPP4i which ensured an increased portal vein GLP-1 concentration [15]. This procedure induced increased vagal firing rates, leading to the stimulation of insulin secretion without increasing systemic concentration of active GLP-1 [15]. Importantly, this regulation would be expected to be impaired in patients with autonomic neuropathy (AN). To investigate this hypothesis further and to avoid the interfering effect of glucose-induced insulin secretion on the regulation glucagon secretion, we subjected T1DM patients characterized by the presence or absence of autonomic diabetic neuropathy to a meal test without or with administration of 50 mg of vildagliptin 30 min before a meal test. Plasma glucose, glucagon, incretins, and pancreatic polypeptide (PP) and selected metabolite concentrations were assessed during the following 180 min.

### Patients and Methods

#### Patients

We included 18 male and female patients with C-peptide negative T1DM, aged 18 - 75 years, insulin treated with multiple daily insulin injections (MDI) or continuous subcutaneous insulin infusion (CSII), diagnosed, according to the current guidelines [27], with confirmed (AN+) or without (AN-) cardiac autonomic neuropathy during autonomic testing realized within the year preceding the enrolment. HbA1c was ≤ 10% and stable (±1%). Females were required to be postmenopausal, surgically sterile, or using a reliable method of contraception. According to the hypothesis, the glucagon responses would be expected to differ markedly according to the presence or absence of neuropathy; therefore, inclusion of 18 individuals should provide the study with sufficient power. It is noteworthy that the study should still be considered only to generate the first evidences to support the hypothesis.

Main exclusion criteria were heart disease (NYHA class III or IV), renal failure (MDRD < 50 mL/min), proliferative retinopathy, liver disease, digestive disease, symptoms of gastroparesis (nausea, vomiting, epigastric pain, and early satiety), any medication that could interfere with the absorption of vildagliptin and ongoing corticosteroid therapy. Patients were allocated into two groups regarding their autonomic neuropathic status (AN- vs. AN+) that were matched for sex, age, and HbA1c, although diabetes duration and daily insulin needs were slightly higher in AN+ than in AN- patients. The characteristics of the 18 patients who completed the study are presented in Table 1.

#### Study design

This was a mechanistic study designed to evaluate whether an impaired autonomic nervous system (neuropathy) would hamper the glucagonostatic effect of the DPP4i, vildagliptin. Patients underwent a meal test (Fortimel Lactose Free containing 29 g carbohydrate, 20 g protein and 7 g lipids) without insulin bolus administration. This procedure ensures a stimulation of glucagon secretion. Patients were overnight fasted while receiving long acting insulin treatment (MDI patients) or basal rate continuous insulin infusion (CSII patients) without insulin

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<th>Table 1. Baseline Characteristics of Patients</th>
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AN+, AN-: patients with or without autonomic diabetic neuropathy, respectively. Data are expressed as mean ± SD, or percent when relevant.
bolus administration in the 3 h preceding the test. In order to avoid major postprandial hyperglycemia, patients had to reach a 0.7 - 1.80 g/L capillary glucose target prior to the meal test. No more than 28 days later, patients underwent the same procedure, except for a single 50 mg vildagliptin oral administration 30 min before the meal test.

Blood samples (tubes containing EDTA 100 nM, aprotinin 2.5 pM and diprotin A 0.1 mM) were obtained at 0, 15, 30, 60, 90, 120 and 180 min. The tubes were immediately centrifuged for 15 min at 4,000 rpm at 4 °C. Plasma was collected and aliquots stored at -80 °C until the concentrations of glucagon, total GIP, active and total GLP-1, PP, insulin, glucose, free fatty acids (FFAs), triglycerides, HDL, LDL, total cholesterol and lactate were assessed. At the end of the test, induced hypoglycemia was corrected under control of the investigator and patient remained in observation for 4 h to monitor unexpected adverse events. The protocol was reviewed and approved by Local Ethic Committee (CPP Sud Ouest and Outre Mer 1) and the French Competent Authority (ANSM). Each patient gave written informed consent prior to participation. This study has been registered on clinicaltrials.gov website, number NCT01452113.

Biochemical analyses

The concentration of plasma glucose was determined by an enzymatic colorimetric procedure (ABX Pentra Glucose HK CP). Plasma insulin concentrations were determined using the human ultrasensitive insulin ELISA kit (Mercodia, Uppsala, Sweden). PP concentration was determined using the human Pancreatic Polypeptide ELISA kit (Millipore, Billerica, USA). The concentrations of plasma FFAs and lactic acid were assessed by colorimetric assays. Lactate was assessed using lac-
tate oxidase which triggers the release of hydrogen peroxide, which reacts with 4-aminophthaldehyde and ESPAS, forming a colored complex in the presence of peroxidase. The intensity of the coloring is proportional to the amount of lactate present in the sample (ABX Pentra Glucose HK CP). FFAs were assessed by enzymatic colorimetric procedure according to the recommendation from the supplier (NEFA C, WAKO, Neus, DE).

Total GLP-1 concentrations were determined with RIA, using a C-terminally directed antiserum (code no. 89390) whereas the active form of the peptide was assessed using a highly specific sandwich ELISA with a sensitivity below 1 pmol/L as described in detail [28]. Total GIP was analyzed using a C-terminally directed antiserum (code no. 80867). Glucagon concentrations in plasma were measured after extraction of plasma with 70% ethanol (vol/vol, final concentration). The antibody employed (code no. 4305) is directed against the C-terminus of the glucagon molecule and therefore mainly measures glucagon of pancreatic origin [29, 30]. Standards were human glucagon and tracer was moniodinated human glucagon (both gifts from NovoNordisk, Bagsvaerd). Sensitivity and detection limit is below 1 pmol/L, intra-assay coefficient of variation is below 6% at 20 - 30 pmol/L, and recovery of standard, added to plasma before extraction, is about 100% when corrected for losses inherent in the plasma extraction procedure.

LDL and HDL cholesterol were assessed by enzymatic colorimetric procedure (ABX Pentra LDL Direct CP and ABX Pentra HDL Direct CP respectively), total cholesterol was analyzed by enzymatic photometric procedure (CHOD-PAP from ABX Pentra Cholesterol CP), and triglycerides (TG) was assessed by an enzymatic colorimetric procedure (ABX Pentra Triglycerides CP).

Plasma DPP4 activity was assessed as previously described [15]. Briefly, DPP4 activity was assessed in 50 µL of plasma incubated with kit reagents for 2 h at 37 °C according to the manufacturers’ recommendations using recombinant DPP4 as a standard, expressed in nanograms per milliliter (DPP4 Glo protease assay; Promega, Madison, WI).

**Autonomic diabetic neuropathy Ewing score**

AN detection was based on Ewing battery (score ranging from 0 to 5) including at least two abnormal heart rate responses in cardio-vagal tests including deep breathing (6 cycles/ min), Valsalva maneuver, 30/15 ratio independently of the presence or not of orthostatic hypotension, changes in blood pressure during standing. Orthostatic hypotension was defined as a reduction in systolic BP of at least 20 mm Hg or diastolic BP at least 10 mm Hg within 3 min standing test (AN+) or without (≤ 1 borderline response to cardio-vagal tests and no orthostatic hypotension) (AN-) cardiac autonomic neuropathy during autonomic testing realized within the year preceding the enrolment. The response to each test was considered as normal (0), borderline (0.5) or impaired (1) according to laboratory normal values for age [31]. According to Ewing score, patients were separated in two groups without (AN-, score < 0.5) or with (AN+, score > 2) confirmed autonomic failure.

**Statistical analyses**

Unless stated otherwise, quantitative variables are expressed as means ± SD and qualitative variables as number (percentage). Statistical analysis was performed using GraphPad Prism statistical software. In bivariate analysis, Chi-square test or Fischer’s exact test when appropriate was used to compare the distribution of qualitative variables between the groups. Means of quantitative variables were compared by Student’s t-test or Mann-Whitney test when appropriate. P < 0.05 was considered significant.

**Results**

**Baseline characteristics of the patients**

At baseline, patients with or without AN were matched for sex, age, insulin treatment, HbA1c, BMI, and all other major diabetic features (Table 1). It is noteworthy that the diabetes duration was slightly higher in the AN+ than in the AN- group.

**Effect of autonomic diabetic neuropathy on plasma peptide and metabolite concentration profiles following a meal test without DPP4i**

We first evaluated the impact of AN on the levels of circulating peptides (PP, glucagon, GLP-1 and GIP) and metabolites (glucose, lactates, FFA and lipids) following a meal test in T1DM patients. The plasma concentration of the PP rapidly increased after the first 15 min following the meal test, but to a lower extent in AN+ patients when compared to AN- controls (Fig. 1A). The plasma glucagon concentrations increased to the same extent in both groups of patients within the first 30 min after the meal test. However, they remained elevated only in the AN+ group of patients while returning towards basal values in AN- patients (Fig. 1B) showing an altered regulation of glucagon in the presence of AN. We then evaluated whether it was linked to an impaired secretion of GLP-1. The plasma concentration of active GLP-1 increased in both groups of patients although to a much lower extent in AN+ patients (Fig. 1C) whereas the plasma concentration profiles of total GLP-1 were similar in both groups (not shown). Thus, the molar ratio of glucagon to active GLP-1 was higher in the AN+ patients than in AN- patients. The plasma GIP concentration profiles were similar in both groups (Fig. 1D).

We then assessed the changes in plasma metabolites. The plasma glucose (Fig. 1E), plasma lactate, FFA, TG, HDL, LDL and cholesterol concentrations were also similar between both groups. The plasma DPP4 activity was similar in the two groups showing a 15-20% decrease during the first 15 - 30 min following the meal test which was maintained for the rest of the observation period (Fig. 1F). There were no differences in the circulating exogenous insulin concentration (not shown). All patients were C-peptide negative, therefore, endogenous insulin did not contribute.
The impact of neuropathy on the control of the concentration profiles of plasma peptides and metabolites in response to a DPP4i was then assessed in the same patients following another meal test less than 28 days later, and a single administration of the inhibitor. The DPP4i did not modify the plasma PP concentration profiles in patients with or without neuropathy (not shown) which remained therefore, similar to what observed in Figure 1A. In contrast, the DPP4i treatment significantly reduced the plasma concentrations of glucagon in AN- patients (Fig. 2A) whereas, this effect was lost for the AN+ patients since the plasma glucagon concentrations remained steadily elevated following the meal test (Fig. 2B). In both AN- and AN+ patients, the plasma concentrations of active GLP-1 were, as expected, increased by the DPP4i treatment (Fig. 2C, D). All other parameters, including glycemia, lactate, FFA, TG, and cholesterol remained unaffected by the acute DPP4i treatment irrespective of the neuropathy (not shown). The lack of metabolic impact of the single administration of DPP4i might be related to the need, in T1DM patients, of either insulin secretion or to a longer exposure to the treatment.

**Discussion**

We here show in T1DM patients that plasma glucagon concentrations following a meal test may be lowered by a DPP4i, obviously via an insulin-independent mechanism, whereas a similar inhibition by DPP4i is not seen in patients with AN indicating that the autonomic nervous system activity is involved in inhibitory activity of the DPP4i. DPP4 inhibition increased similarly the levels of intact GLP-1 in patients with as well as without neuropathy. Therefore, given that elevated levels of intact GLP-1 are responsible for the normal inhibition of glucagon secretion, our data also suggest that AN induces a state of glucagon unresponsiveness representing a form of GLP-1 resistance.

Patients with both T1DM and T2DM have impaired regulation of glucagon secretion which may be responsible for excessive hepatic glucose production [32, 33]. In addition, unlike healthy humans, patients with T1DM do not respond to hypoglycemia with increased glucagon secretion, and this is thought to be of importance for the high frequency of hypoglycemic attacks during insulin therapy in these patients. The mechanisms involved are yet unclear but could be related, to some degree, to AN which often affects these patients [34, 35]. We provide here evidence to support this hypothesis by showing that AN is associated with an exaggerated glucagon response to a meal test in insulin-deprived T1DM patients. The mechanism could not be related to disturbed insulin action since all patients, with or without neuropathy, were C-peptide negative. These observations may therefore be consistent with a role for the autonomic nervous system in the control of glucagon secretion [36, 37]. Previous work in rodents showed that vagotomy [38] as well as ventromedial hypothalamic lesions [39] hampers the regulation of glucagon secretion following
feeding in rats. We here further showed that the meal responses of PP were lower in patients with neuropathy in agreement with the notion that activity in efferent cholinergic neurons of the parasympathetic nervous system is responsible for a considerable part of PP response to a meal. Acetylcholine, the parasympathetic transmitter has been shown to reduce glucagon secretion on isolated perfused pancreas [40]. The exaggerated glucagon response in the neuropathy patients might therefore be due to an impaired cholinergic, parasympathetic inhibition of glucagon secretion. A 80% reduction in the number of nerve endings in direct contact with alpha-cells was noted type 1 diabetes BB rats [41] which could be responsible for an impaired glucagon regulation as well as reduced PP secretion. However, acetylcholine activates glucagon secretion from isolated pancreatic alpha cells [42, 43], indicating that glucagonostatic effect of acetylcholine on the whole pancreas is due to a more complex mechanism, perhaps involving inhibition of somatostatin secretion [37, 44]. Our data are in agreement with this conclusion since the plasma glucagon to PP concentration ratio increased during the meal test in AN+ patients while it remained low and steady under control in AN- patients. Furthermore, the glucose unresponsiveness of the patients with AN could be due to a state of GLP-1 resistance. The impaired glucagonostatic action of the active GLP-1 was not due to an impaired secretion of the peptide since total plasma GLP-1 concentration was normal. Hence, a tight relationship is suggested between the autonomic parasympathetic nerve activity and the action of endogenous GLP-1 on glucagon concentration. Our conclusion is also supported by recent data from the literature showing that the deletion of the GLP-1 receptor specifically in the β cells did not impair oral glucose induced insulin secretion in mice [45] further supporting the importance of GLP-1 receptors expressed in other than beta cells. Therefore, as we previously suggested, endogenous GLP-1 may control pancreatic endocrine secretions mainly through neural mechanisms [10, 46-48], whereas GLP-1 receptor agonists reaching high circulating concentrations of active agonist would control glycemia through a direct effect on β cells [49].

Interestingly, neuropathy also appeared to reduce meal test-induced active GLP-1 plasma concentration suggesting that GLP-1 homeostasis is under neural regulation. The impaired GLP-1 response may contribute to the impaired regulation of glucagon secretion observed in patients with AN. Possibly an impaired function of the autonomic nervous system may result in impaired GLP-1 secretion, perhaps by influencing gastrointestinal motility [50-52]. However, our data show that the total GLP-1 concentration was not dramatically affected by the AN, most likely ruling out impaired neural regulation of GLP-1 secretion. Furthermore, no change in DPP4 activity was observed. The acute changes observed in plasma glucagon and GLP-1 concentrations did not affect the overall glycemic profiles following the meal test suggesting that chronic long term rather than acute changes in plasma GLP-1 and glucagon concentrations are required to improve the glycemic control.

Since our data might suggest that lower concentrations of active GLP-1 could explain the exaggerated glucagon response in AN, we treated the AN+ T1DM patients with a DPP4i to enhance GLP-1 action. Indeed, the DPP4i efficiently increased the plasma concentrations of active GLP-1 but, however, failed to improve the glucagon concentration profiles during the meal test in AN+ patient. Our findings therefore suggest that AN increases meal-induced glucagon secretion through a mechanism preventing the glucagonostatic effect of enteric GLP-1 on the gut to alpha cell axis in T1DM patients.

In T1DM patients without AN, our data show that the administration of a DPP4i, to enhance the physiological action of gut-released GLP-1, inhibits glucagon secretion by an insulin-independent mechanism with the additional information that the mechanism requires an intact autonomic nervous system. Also GLP-1 agonists inhibit glucagon secretion via insulin-independent mechanisms [5, 53]. Our data are also in agreement with previous observations with vildagliptin in patients with T1DM [3, 54].

Although we cannot rule out that the GLP-1 receptor present at the surface of the alpha cells could be responsible for the direct glucagonostatic effect of the GLP-1 secreted by the gut, the demonstration that vagotomy abolishes it supports the role of a neural mechanism [22]. Electrophysiological recording of the vagus nerve discharge in the presence of GLP-1 demonstrates the direct effect of the peptide [16, 55]. Long vago-vagal reflexes in turn would regulate pancreatic and extra pancreatic functions such as insulin secretion [13], whole body glucose turnover [46, 56-59] and liver metabolism [60] participating in general gut-to-brain-to-periphery regulation of metabolism. Impaired gastric emptying in patients with AN could have contributed to the impaired glucagon secretion, but in our study patients with symptomatic gastroparesis were excluded, although gastric emptying was not measured in all the patients. Furthermore, the glycemic peak and the maximal early plasma glucagon concentrations reached following the meal test were similar in patients with or without neuropathy showing that the overall meal has been most likely similarly absorbed. A recent study in patients with T2DM demonstrated no effect of vildagliptin on the rate of gastric emptying or meal glucose appearance [61] suggesting that this issue was not at play in our study.

Importantly and to the best of our knowledge, the integrity of the autonomic nervous system has never been considered as a criterion to predict the responsiveness of alpha cells to the glucagonostatic effect of a GLP-1-based therapy. This criterion could define responders versus non-responder patients. In addition to DPP4i responsiveness, our conclusion could also be true for GLP-1 receptor agonists which would require an integrated autonomic nervous system to efficiently control glucagon secretion and hence glycemia. Although, whether high circulating GLP-1 concentration, as obtained using GLP-1 receptor agonist, is sufficient to activate the gut brain to pancreas axis remains to be determined.

Altogether, we here show that the AN, which characterizes many T1DM and T2DM patients [34, 35], impairs the gut to alpha cell axis regarding GLP-1 regulated glucagon secretion. Consequently, the autonomic diabetic neuropathy could hamper the regulation of glycemia by DPP4i. This would be of major importance for the treatment of T2DM since both insulin and glucagon secretion would be impaired by AN.
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