Is It Relevant to Screen Subclinical Cushing’s Syndrome in Patients With Type 2 Diabetes Mellitus?

Manel Jemel Hadiji, Imen Ksir, Emna Haouat, Henda Kammoun Jamoussi, Leila Ben Salem Hachmi

Abstract

Background: Subclinical Cushing’s syndrome (SCS) is defined as autonomous cortisol secretion in patients devoid of specific clinical symptoms of hypercortisolism, as in the classic CS. However, sustained exposure to chronic slightly elevated cortisol concentrations may result in some classical metabolic complications of CS such as impaired glucose tolerance and diabetes. Currently, the frequency of SCS is widely variable. We have conducted a cross-sectional study to prospectively evaluate the prevalence of SCS among type 2 diabetic (T2D) patients with poor control, and to determine whether systematic screening for SCS in T2D patients is worthwhile.

Methods: It was a cross-sectional study including 221 T2D patients referred to the National Institute of Nutrition of Tunis for poor glycemic control (HbA1c ≥ 8%). The first screening step of SCS was the 1-mg overnight dexamethasone suppression test (ODST) using a revised criterion for cortisol suppression. In the second confirmatory step, patients with abnormal ODST underwent a 48-h, 2-mg low-dose dexamethasone suppression test (LDDST) to confirm the diagnosis. The cut-off for cortisol suppression was 50 nmol/L (1.8 µmol/dL) in the two tests.

Results: Thirteen patients (5.9%) failed to suppress cortisol levels less than the cut-off after ODST. SCS was confirmed by LDDST in one patient among them (0.45%). The autonomous cortisol secretion was related to a pituitary adenoma. Our study revealed that the frequency of SCS of 0.45% did not allow performing an analytical study in order to identify predictive factors of SCS among T2D patients.

Conclusion: SCS is rare among T2D patients. Systematic screening of SCS in T2D patients with poor glycemic control is not worthwhile. The screening should be performed only in patients with a particular clinical and/or biological context.

Keywords: Type 2 diabetes; Cushing’s syndrome; Dexamethasone suppression test; Obesity; Metabolic syndrome
min) or hepatic failure, depression and alcoholism were excluded.

**Methods**

The selected patient underwent a 1-mg overnight dexamethasone (DXM) suppression test (ODST) as the first step screening test. They took 1 mg of DXM at 23:00 and the sample blood test was collected on the next morning at 08:00 am for measurement of serum level.

The cut-off for cortisol suppression was 50 nmol/L (1.8 µg/dL). The patients who had cortisol less than 50 nmol/L (1.8 µg/dL) were considered normal and did not need further exploration. If the cortisol level was above this cut-off, the patient was considered abnormal and underwent a 2-mg low-dose dexamethasone suppression test (LDDST) with the same cortisol cut-off. He took a 0.5 mg of DXM at 06:00, 12:00, 18:00 and 24:00 for two consecutive days. At the third day, a blood sample test was used for cortisol measurement [1, 16]. In the case of a positive diagnosis of CS with a lack of cortisol suppression in the LDDST, patients were hospitalized for etiological exploration and specific management. In addition, adrenocorticotropic hormone was measured. For the 8-mg DST, suppression was present when there was a decrease in cortisol concentration by more than 50% from baseline [17].

The clinical and biological characteristics of the patients were analyzed. Body mass index (BMI) values, defined by the weight ratio (kg) on the square of the size (in meters), was interpreted in accordance with the classification adopted by the World Health Organization in 1998 [18]. Waist circumference has been interpreted according to the recommendations of the International Diabetes Federation (IDF): the android fat distribution was defined by a waist circumference greater than 94 cm for men and 80 cm for women [19]. Patients were considered hypertensive if they were: 1) already known hypertensive and treated; and 2) having a blood pressure ≥ 140/90 mm Hg confirmed by at least two outpatient measures.

For hypertensive patients, blood pressure was considered well controlled when systolic blood pressure was < 140 mm Hg and diastolic blood pressure was < 80 mm Hg (American Diabetes Association goals 2014) [20]. Low-density lipoprotein-cholesterol (LDL-C) was calculated according to the FRIEDWALD formula (for triglycerides (TG) < 4.4 mmol/L): LDL-C (g/L) = TG (mmol/L)/2.2 × 0.387. High cholesterol was defined as total cholesterol ≥ 5.2 mmol/L or by taking a statin. Hypertriglyceridemia was defined as a TG level ≥ 1.7 mmol/L or by taking fibrates. Creatinine clearance was evaluated using the Cockcroft and Gault formula: creatinine clearance (mL/min) = (140 - age) × weight (kg)/creatinine (µmol/L) × k; in women: k = 1.04; in human: k = 1.25. Hypokalemia was defined as serum potassium < 3.5 mmol/L.

**Statistical analysis**

Statistical analyses were performed with SPSS software, version 13.0.

**Results**

A total of 221 patients were included in the study. The mean age ± SD of the patients was 58.7 ± 8.78 years. Overweight and obesity were found in 34% and 47%, respectively. An android fat distribution was present in 76% of patients. The mean duration of diabetes was 10.84 ± 6.55 years. Hypertension was present in 35.7%, with poor control in 34.2%. In the study, our patients had hypercholesterolemia and hypertriglyceridemia in 63.8% and 47.1%, respectively. The mean baseline HbA1c ± SD was 11.5 ± 2.4.

Table 1. Comparison of Characteristics of Patients According to ODST

<table>
<thead>
<tr>
<th></th>
<th>Cortisol ≥ 50 nmol/L</th>
<th>Cortisol &lt; 50 nmol/L</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>Male (n)</td>
<td>2</td>
<td>66</td>
<td>0.18</td>
</tr>
<tr>
<td>Female (n)</td>
<td>11</td>
<td>142</td>
<td>0.18</td>
</tr>
<tr>
<td>Age (years)</td>
<td>59 ± 8</td>
<td>58.6 ± 8.8</td>
<td>0.9</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>33.65 ± 6.5</td>
<td>29.9 ± 5.3</td>
<td>0.016</td>
</tr>
<tr>
<td>Waist circumference (cm)</td>
<td>104.46 ± 13.4</td>
<td>101.5 ± 11.74</td>
<td>0.39</td>
</tr>
<tr>
<td>Type 2 diabetes duration (years)</td>
<td>9.38 ± 4.2</td>
<td>10.9 ± 6.7</td>
<td>0.41</td>
</tr>
<tr>
<td>Triglycerides (mmol/L)</td>
<td>1.73 ± 0.62</td>
<td>1.84 ± 1.06</td>
<td>0.71</td>
</tr>
<tr>
<td>Total cholesterol (mmol/L)</td>
<td>4.46 ± 1.55</td>
<td>4.98 ± 1.18</td>
<td>0.26</td>
</tr>
<tr>
<td>LDL-cholesterol (mmol/L)</td>
<td>1.08 ± 0.51</td>
<td>1.16 ± 0.37</td>
<td>0.42</td>
</tr>
<tr>
<td>HDL-cholesterol (mmol/L)</td>
<td>1.03 ± 0.28</td>
<td>1.14 ± 0.28</td>
<td>0.19</td>
</tr>
<tr>
<td>HbA1c (%)</td>
<td>11.5 ± 2.4</td>
<td>10.8 ± 1.8</td>
<td>0.22</td>
</tr>
<tr>
<td>1-mg ODST (nmol/L)</td>
<td>91.8 ± 76.1</td>
<td>26.26 ± 8.95</td>
<td>0.009</td>
</tr>
</tbody>
</table>

BMI: body mass index; ODST: 1-mg overnight dexamethasone suppression test; LDL: low-density lipoprotein; HDL: high-density lipoprotein.
after diabetes diagnosis. As non-specific signs of CS, facial erythrosis was noted in 23.1%, Acanthosis nigricans in 16.3%, and buffalo neck in 11.8% of cases. The association of excess weight with facial erythrosis or buffalo neck was noted in 19% and 11.3%, respectively.

Thirteen (5.9%) out of 221 patients failed to suppress cortisol on ODST and had a mean cortisol concentration after this test of 91.87 ± 76.15 nmol/L (Table 1). There was a significant difference in BMI between DST suppressors and non-suppressors, but no significant difference in age, sex, or other metabolic parameters such as glucose, HbA1c, and cholesterol. All non-suppressors were further evaluated by a 2-mg LDDST. Out of these 13, one patient (0.45%) had a cortisol level > 50 nmol/L after LDDST (71.87 nmol/L), but he was a suppressor on 8-mg DST (Fig. 1). For this patient, the ACTH was at 17 ng/L and the patient had a pituitary lesion in magnetic resonance imaging (right anterior pituitary lesion 1.3 mm hyposignal T1, in hypersignal T2, taking strongly the contrast with the healthy pituitary, Figs. 2 and 3). As the clinical and biological presentation was not severe, we repeat a second MRI after 6 months. The second MRI showed an increase in the size of the pituitary lesion measuring 2 mm. The patient was then referred for neurosurgery for microadenoma.

**Figure 1.** Results of the screening strategy for subclinical Cushing’s syndrome in 221 type 2 diabetic patients with poor glycemic control. DST: dexamethasone suppression test.

**Figure 2.** Pituitary lesion in hyposignal T1 with intense enhancement after injection of gadolinium.

**Figure 3.** Pituitary lesion in hypersignal T2.
resection.

Discussion

Type 2 diabetes is a condition of chronic hyperglycemia, the main physiopathological mechanism of which is insulin resistance, followed by progressive insulopenia [21, 22]. This insulin resistance during type 2 diabetes results in a clinical expression identical to that found during SCS, making it even less obvious the diagnosis of an SCS associated with diabetes [12, 23]. Thus, we performed a systematic screening of SCS in T2D patients with chronic poor glycemic control. Of the 221 patients in our series, only one patient had an SCS (a prevalence of 0.45% of SCS in poor controlled T2D patients). Several other studies have been conducted since 1996 to determine the prevalence of SCS in populations of T2D subjects. The number of T2D patients included in these studies ranged from 77 to 993; the prevalence of SCS among reported T2D patients ranged from 0% to 9.4% [10, 13, 14, 24-33]. Differences in the results of all these studies can be explained mainly by the methodological differences: inclusion criteria, screening methods and their cut-off values, and even the hormonal diagnostic criteria of SCS since there is no definition of the SCS. Table 2 summarizes the main studies on SCS testing in T2D patients and their methodological differences.

<table>
<thead>
<tr>
<th>Studies</th>
<th>Number of patients (n)</th>
<th>Inclusion criteria</th>
<th>Screening method (and cut-off value)</th>
<th>Prevalence of SCS, n (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Chiodini et al [14]</td>
<td>294</td>
<td>T2D Age at diagnosis of diabetes &gt; 30 years 19 kg/m² ≤ BMI &lt; 50 kg/m²</td>
<td>1-mg DST (50 nmol/L)</td>
<td>30 (9.4%)</td>
</tr>
<tr>
<td>Murakami et al [31]</td>
<td>90</td>
<td>T2D Age &gt; 20 years 16 kg/m² ≤ BMI ≤ 25 kg/m²</td>
<td>Midnight serum cortisol (69 nmol/L) + 1-mg DST (83 nmol/L)</td>
<td>8 (8.9%)</td>
</tr>
<tr>
<td>Leibowitz et al [28]</td>
<td>90</td>
<td>T2D HbA1c &gt; 9% BMI &gt; 25 kg/m²</td>
<td>1-mg DST (140 nmol/L)</td>
<td>3 (3.3%)</td>
</tr>
<tr>
<td>Caetano et al [25]</td>
<td>103</td>
<td>T2D Age &gt; 20 years BMI &gt; 25 kg/m²</td>
<td>Midnight salivary cortisol (7 nmol/L) + 1-mg DST (50 nmol/L)</td>
<td>3 (2.9%)</td>
</tr>
<tr>
<td>Taniguchi et al [10]</td>
<td>77</td>
<td>T2D</td>
<td>Midnight serum cortisol (138 nmol/L)</td>
<td>2 (2.6%)</td>
</tr>
<tr>
<td>Catargi et al [26]</td>
<td>200</td>
<td>T2D BMI &gt; 25 kg/m² HbA1c &gt; 8%</td>
<td>1-mg DST (58 nmol/L)</td>
<td>4 (2%)</td>
</tr>
<tr>
<td>Reimondo et al [33]</td>
<td>100</td>
<td>T2D recently diagnosed</td>
<td>1-mg DST (110 nmol/L)</td>
<td>1 (1%)</td>
</tr>
<tr>
<td>Terzolo et al [13]</td>
<td>813</td>
<td>T2D BMI &gt; 25 kg/m² Age between 18 and 70 years</td>
<td>1-mg DST (138 nmol/L)</td>
<td>6 (0.7%)</td>
</tr>
<tr>
<td>Liu et al [29]</td>
<td>154</td>
<td>Elderly male subjects with T2D</td>
<td>Midnight salivary cortisol (4.3 nmol/L)</td>
<td>0 (0%)</td>
</tr>
<tr>
<td>Budyal et al [24]</td>
<td>993</td>
<td>Outpatients DT2</td>
<td>1-mg DST (50 nmol/L)</td>
<td>0 (0%)</td>
</tr>
<tr>
<td>Newsome et al [32]</td>
<td>171</td>
<td>T2D BMI &gt; 25 kg/m² Age 18 - 80 years</td>
<td>1-mg DST (50 nmol/L)</td>
<td>0 (0%)</td>
</tr>
<tr>
<td>Gagliardi et al [27]</td>
<td>101</td>
<td>T2D Age 40 - 75 years HbA1c &gt; 8% BMI &gt; 25 kg/m² Waist size &gt; 80 cm (woman) and &gt; 94 cm (man)</td>
<td>Midnight salivary cortisol (13 nmol/L)</td>
<td>0 (0%)</td>
</tr>
<tr>
<td>Mullan et al [30]</td>
<td>201</td>
<td>T2D HbA1c &gt; 7% BMI &gt; 25 kg/m² Blood pressure &gt; 140/90 mm Hg</td>
<td>Midnight salivary cortisol (10 nmol/L)</td>
<td>0 (0%)</td>
</tr>
</tbody>
</table>
study, the present study showed that cortisol suppression to O DST with the cut point of 1.8 µg/dL was false positive in 13 (5.8%) patients and there is a significant correlation with BMI. Other potential confounders including stress, poor controlled diabetes, and the activators of cytochrome P-450 3A4 system such as statins should be considered. These results emphasized that high false positivity of O DST should be considered in the clinical practice and additional tools such as the 24-h urinary free cortisol or the late-night salivary cortisol test must be employed to confirm the diagnosis of CS.

Fixing the cortisol cut-off value at 50 nmol/L (which improves the sensitivity of the test), and eliminating situations that may induce pseudo-CS, the specificity of the 1-mg DST in our series was 94.5%. In the other studies carried out in T2D patients, the reported specificities of the 1-mg DST range between 78% and 97.7%. The comparison of our findings with those of the studies is difficult because, for the majority of these studies, the cut-off values for the DST as well as the SCS confirmation criteria were different from ours [14] (Table 2).

In our study, only one patient finally presented an SCS confirmed by the 2-day 2-mg DST. Given this result (a patient with SCS and 220 patients without SCS), we could not rely on this isolated case to perform an analytical study to identify predictive factors for SCS.

Screening for SCS in T2D patients has three foundations. A higher incidence of SCS in T2D patients compared with non-diabetic patients. Indeed, Chiodini et al [14] showed in his study that the prevalence of SCIC in T2D patients was significantly higher than in non-diabetics who were matched for age and BMI (9.4% vs. 2.1%; P = 0.004). This frequency could reach 42% in T2D patients [15].

SCS is an independent and important cardiovascular risk factor to manage to reduce the morbidity and mortality of patients with T2D patients who are already at high cardiovascular risk. Several studies have shown an increased cardiovascular risk in patients with SCS. This risk is related both to the metabolic consequences and to a direct effect of excess cortisol on the vascular walls [9, 35-38].

The management of SCS would lead to an improvement in metabolic disorders. Indeed, Tachmanova et al [39] reported that the SCS treatment improves both glycemic control and hypertension and also allows weight loss. Taniguchi et al [10] clearly demonstrated an improvement in insulin sensitivity after treatment of SCS pituitary with a hyperinsulinemic eu-glycemic clamp, a reference technique for measuring insulin sensitivity [40-42]. Both patients had a marked improvement in glycemic control after treatment. In contrast, no randomized controlled trials demonstrating the impact of SCS on metabolic profile and cardiovascular mortality have been published to date [7, 15].

Based on a frequency argument, and knowing that the prevalence of SCS in the general population is estimated to be between 0.2% and 2%, our results do not suggest a particular interest in systematic SCS screening in poorly controlled T2D patients, in any case no more than in the general population. Thus, given the low incidence of SCS in poorly controlled T2D patients, and the absence of studies demonstrating the benefit of SCS treatment on metabolic profile and cardiovascular morbidity and mortality, routine SCS screening in T2D patients does not seem relevant. Rather, we recommend: 1) a physical examination of diabetic patients in particular looking for signs of hypercatabolism; specific signs of the franc CS, and particular attention to certain biological parameters (e.g. hypokalemia associated with hypertension) and certain co-morbidities (resistant hypertension, “idiopathic” osteoporosis). These situations represent grounds for CS research even outside of diabetes.

In the literature, many other authors do not recommend SCS testing in all T2D patients. Indeed, the majority of these authors recommend rather a targeted screening based essentially on the clinicobiological characteristics of the patients detected in their series.

For example, Terzolo et al [13] considered that the motivating factors for SCS screening were poorly controlled T2D patients, resistant hypertension, age of diagnosis of diabetes below 50 years, and lack of family history of type 2 diabetes. Chiodini et al, who found the highest prevalence of SCS among T2D patients [14], showed that SCS was significantly associated with insulin-treated diabetes, poorly controlled and coexisting with hypertension or dyslipidemia. Six years after the publication of this study, they suggested that screening would be beneficial in patients with poorly controlled T2D patients despite appropriate treatment and adherence, especially those with associated osteoporosis or hypertension [7].

Finally, in a review article published in 2015 entitled “Screening for Cushing’s syndrome: is it worthwhile [43]?” Shimon considers that the routine screening of SCS in T2D patients is not warranted. It should be considered only in young patients with resistant T2D patients or resistant HTA. The author did not specify the age limit and the definition of resistant diabetes.

Conclusions

SCS is rare among T2D patients. There is no benefit in routinely screening SCS for T2D patients with chronic poor glycemic control. This screening should be targeted and guided by the clinical and biological context.

References

5. Lindholm J, Juul S, Jorgensen JO, Astrup J, Bjørre P,


36. Giordano R, Marinazzo E, Berardelli R, Picu A, Mac- 
cario M, Ghigo E, Arvat E. Long-term morphological, 
hormonal, and clinical follow-up in a single unit on 118 
2010;162(4):779-785.
37. Siren J, Tervahartiala P, Sivula A, Haapiainen R. Natural 
course of adrenal incidentalomas: seven-year follow-up 
of the metabolic syndrome? J Clin Endocrinol Metab. 
39. Tauchmanova L, Rossi R, Biondi B, Pulcrano M, Nuzzo 
V, Palmieri EA, Fazio S, et al. Patients with subclinical 
Cushing’s syndrome due to adrenal adenoma have in-
creased cardiovascular risk. J Clin Endocrinol Metab. 
2002;87(11):4872-4878.
40. DeFronzo RA, Tobin JD, Andres R. Glucose clamp tech-
nique: a method for quantifying insulin secretion and re-
41. Rabasa-Lhoret R, Laville M. [How to measure insu-
lin sensitivity in clinical practice?]. Diabetes Metab. 
42. Taniguchi T, Okamoto M, Ueno H, Tanaka J, Okamoto M, 
Hamasaki A, Yamada Y, et al. Use of the insulin sensitiv-
ity index obtained from oral glucose tolerance test in Jap-
144.
43. Shimon I. Screening for Cushing’s syndrome: is it worth-